



Host-response based endotyping and prognostication of patients with COVID19 – the Greek experience

E. J. Giamarellos-Bourboulis, MD, PhD

Professor of Internal Medicine

4th Department of Internal Medicine
Director MSc Infectious Diseases

National & Kapodistrian University of Athens, Medical School, Greece

President: European Shock Society
Chairman: European Sepsis Alliance

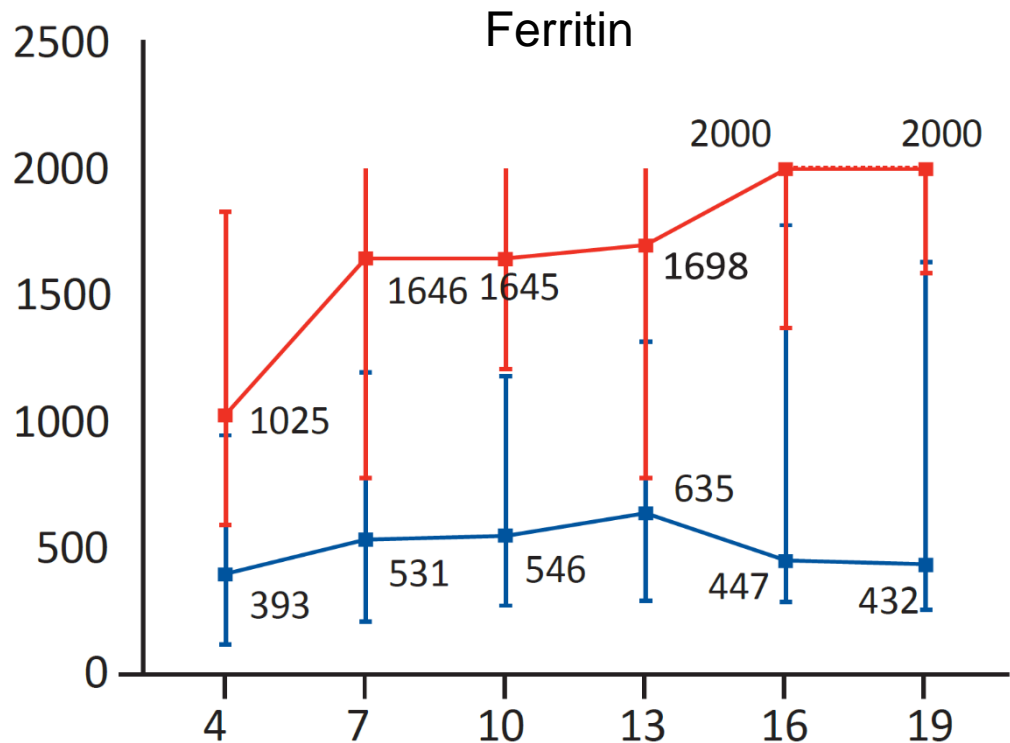
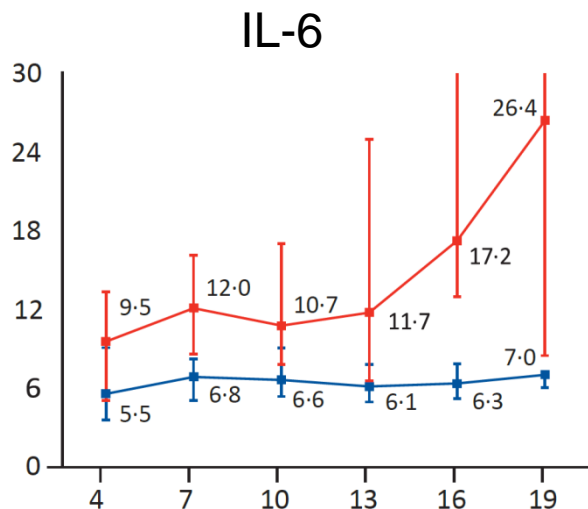


CONFLICT OF INTEREST DISCLOSURE

- Honoraria (paid to the University of Athens) from Abbott CH, Brahms ThermoFisher GmbH Germany, GSK, Inflammatix Inc and Sobi
- Consultant for Fab'nTech, InflaRx GmbH, UCB and Xbiotech Inc
- Independent educational grants (paid to the University of Athens) from AbbVie USA, InflaRx GmbH, Novartis, UCB
- Independent educational grants (paid to the Hellenic Institute for the Study of Sepsis) from Abbott CH, BioMérieux France, MSD, Inflammatix Inc Sobi, ThermoFisher Brahms GmbH, Xbiotech Inc
- Funding by the Horizon 2020 ITN European Sepsis Academy (granted to the University of Athens) and by the Horizon 2020 ImmunoSep and RISKinCOVID (granted to the Hellenic Institute for the Study of Sepsis)

IMMUNE RESPONSE IN COVID-19: TRAITS OF MACROPHAGE ACTIVATION

(Zhou F, et al. *Lancet* 2020; 395: 1-54-1062)



Survivors (n=137)

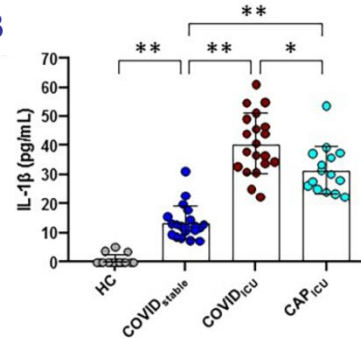
Non-survivors (n=54)

IMMUNE RESPONSES IN COVID-19: TH1 TO TH2 IMBALANCE

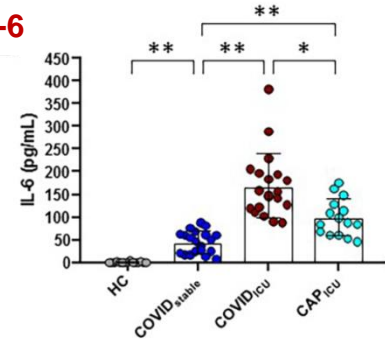
(McElvaney OJ, et al. *Am J Resp Crit Care Med* 2020; 202: 812-81)

- Healthy (HC, n=20)
- COVID_{stable} (n=20)
- COVID_{ICU} (n=20)
- CAP: community-acquired pneumonia (n=20)

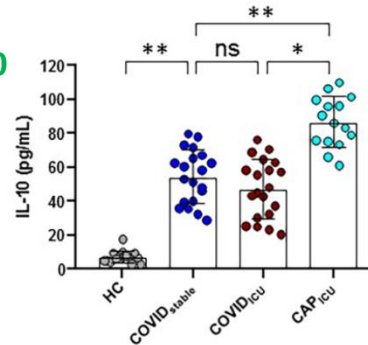
IL-1 β



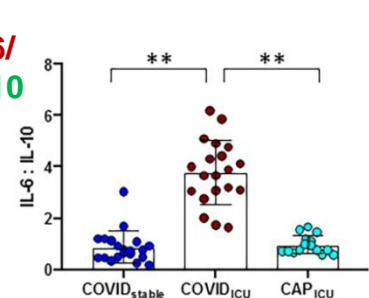
IL-6



IL-10



IL-6/
IL-10



*p<0.05
**p<0.01

THE SEPSIS PARADIGM OF HETEROGENEITY

EBioMedicine 6 (2018) 114–125



Contents lists available at ScienceDirect

EBioMedicine

journal homepage: www.elsevier.com/locate/ebio



Research Paper

A Transcriptomic Biomarker to Quantify Systemic Inflammation in Sepsis – A Prospective Multicenter Phase II Diagnostic Study



Michael Bauer^{a,b,*}, Evangelos J. Giamarellos-Bourboulis^{b,c}, Andreas Kortgen^{a,b}, Eva Möller^d, Karen Felsmann^d, Jean Marc Cavailhon^e, Orlando Guntinas-Lichius^f, Olivier Rutschmann^g, Andriy Ruryk^h, Matthias Kohl^h, Britta Wlotzkaⁱ, Stefan Rufswurm^a, John C. Marshall^j, Konrad Reinhart^{a,b}

^a Department of Anesthesiology and Intensive Care Medicine, Jena University Hospital, Erlanger Allee 101, 07546 Jena, Germany

^b Center for Sepsis Control & Care, Jena University Hospital, Erlanger Allee 101, 07546 Jena, Germany

^c 6th Department of Internal Medicine, University of Athens Medical School, 7 Soranou Efessiou, 11527 Athens, Greece

^d Andriy-Jena AG Germany, Sepsis diagnostics rapid group (previously SIRS-Lab GmbH Jena), Konrad-Zuse-Straße 1, 07745 Jena, Germany

^e Unit of Critical Care Medicine, Hôpital Pasteur, 73015 Paris, France

^f Department of Otolaryngology and the Institute of Rhinology and Pediatry, Jena University Hospital, Lotharstraße 2, 07740 Jena, Germany

^g Division of Emergency Medicine, Department of Community, Primary Care and Emergency Medicine, Geneva University Hospital and Faculty of Medicine, 7, rue Gabrielle Perret Cassin, 1211 Geneva 14, Switzerland

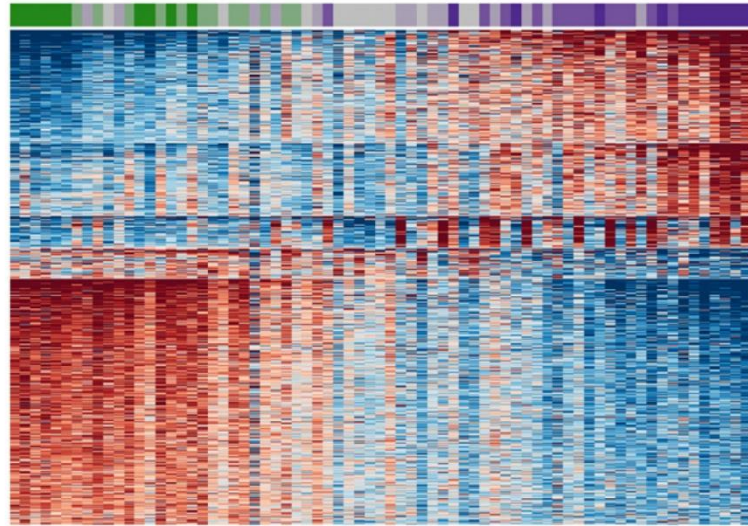
^h Faculty of Medical and Life Sciences, Fortwangen University, Jakob-Klein-Str. 17, 78054 Villingen-Schwenningen, Germany

ⁱ State Enterprise Corporation of Therapia, Malzerhofstraße 12, 39100 Bfz, Germany

^j University of Toronto, St. Michael's Hospital, 37 Bond Street, Room 4104, Toronto, Ontario M5B 1A8, Canada

Low-grade
inflammation

High-grade
inflammation



- No signs of inflammation
- Sterile local infl.
- Local infection w/o systemic infl.
- Systemic infl. w/o infection
- Local infection with systemic infl.
- BSI with signs of systemic infl.



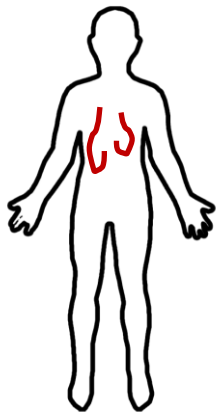
OPEN

Early treatment of COVID-19 with anakinra guided by soluble urokinase plasminogen receptor plasma levels: a double-blind, randomized controlled phase 3 trial

Evdoxia Kyriazopoulou¹, Garyfallia Poulakou², Haralampos Milionis³, Simeon Metallidis⁴, Georgios Adamis⁵, Konstantinos Tsiakos⁶, Archontoula Fragkou⁷, Aggeliki Rapti⁶, Christina Damoulari¹, Massimo Fantoni⁸, Ioannis Kalomenidis⁹, Georgios Chrysos¹⁰, Andrea Angheben¹¹, Ilias Kainis¹², Zoi Alexiou¹³, Francesco Castelli¹⁴, Francesco Saverio Serino¹⁵, Maria Tsilika¹, Petros Bakakos¹⁶, Emanuele Nicastrì¹⁷, Vassiliki Tzavara¹⁸, Evangelos Kostis¹⁹, Lorenzo Dagna²⁰, Panagiotis Koufargyris¹, Katerina Dimakou²¹, Spyridon Savvanis⁷, Glykeria Tzatzagou²², Maria Chini²³, Giulio Cavalli²⁰, Matteo Bassetti²⁴, Konstantina Katrini¹, Vasileios Kotsis²⁵, George Tsoukalas²⁶, Carlo Selmi²⁷, Ioannis Bliziotis²⁸, Michael Samarkos²⁹, Michael Doulas³⁰, Sofia Ktena¹, Aikaterini Masgala³¹, Ilias Papanikolaou³², Maria Kosmidou³, Dimitra-Melia Myrodi², Aikaterini Argyraki³³, Chiara Simona Cardellino¹¹, Katerina Koliakou³⁴, Eleni-Ioanna Katsigianni³⁴, Vassiliki Rapti², Efthymia Giannitsioti¹⁰, Antonella Cingolani⁸, Styliani Micha³⁴, Karolina Akinosoglou³⁵, Orestis Liatsis-Douvitsas³⁴, Styliani Symbardi³⁶, Nikolaos Gatselis³⁷, Maria Mouktaroudi^{1,34}, Giuseppe Ippolito¹⁷, Eleni Florou³⁴, Antigone Kotsaki¹, Mihai G. Netea^{38,39}, Jesper Eugen-Olsen⁴⁰, Miltiades Kyprianou³⁴, Periklis Panagopoulos⁴¹, George N. Dalekos³⁷ and Evangelos J. Giamarellos-Bourboulis^{1,34} ✉

suPAR-GUIDED ANAKINRA TREATMENT FOR VALIDATION OF THE RISK AND
EARLY MANGEMENT OF SEVERE RESPIRATORY FAILURE BY COVID-19

THE SAVE STRATEGY



STOP
IL-1 α
IL-1 β



PREVENT
Unfavorable outcome

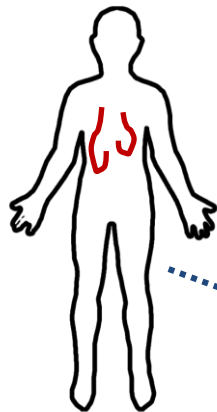
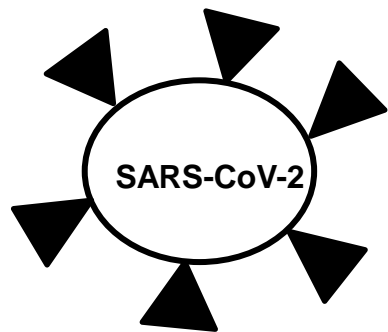


Early identification of risk

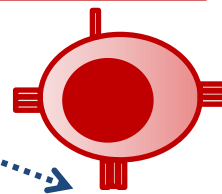
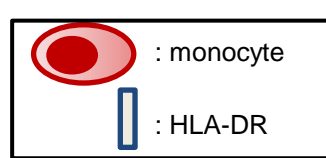
- COVID-19 pneumonia
- Hospitalization
- pO₂/FiO₂: 150-400
- Oxygen mask/nasal oxygen
- suPAR \geq 6 ng/ml

Anakinra

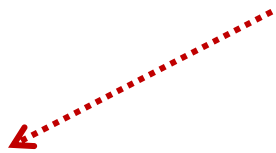
- Recombinant human receptor antagonist
- Block the action of IL-1 α and IL-1 β



Infiltrates
↑CRP
↑D-dimers
↑AST/ALT

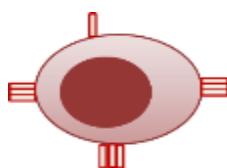
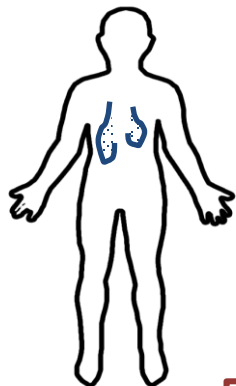


Vivid antigen-
presentation



Macrophage activation: IL-1 β (25%)

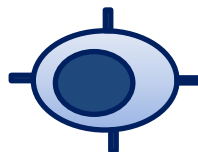
Immune dysregulation: IL-6 (75%)



↑↑↑CRP/ferritin/TGs
↑↑D-dimers
↑↑AST/ALT

↑↑TNF α
↑↑IL-1 β
↑↑IL-6

Moderate antigen-
presentation



↑↑CRP
↑↑D-dimers
↑↑AST/ALT

Weak antigen-
presentation

↑↑TNF α
↑↑IL-6

↓↓ CD4-/CD8-/T17-
lymphocytes
↓ B-lymphocytes, ↓IgGs
↓↓ NKT-/NK-cells

HOW COULD SEVERE COVID-19 BE CLASSIFIED?

Hyper-inflammation*



ARG1, LCN2, LTF, OLFM4, HLA-DMB

Adaptive immunity*



YKT6, PDE4B, TWISTNB, BTN2A2, ZBTB33, PSMB9, CAMK4, TMEM19, SLC12A7, TP53BP1, PLEKHO1, SLC25A22, FRS2, GADD45A, CD24, S100A12, STX1A

Hyper-coagulation*



KCNMB4, CRISP2, HTRA1, PPL, RHBDF2, ZCCHC4, YKT6, DDX6, SENP5, RAPGEF1, DTX2, RELB

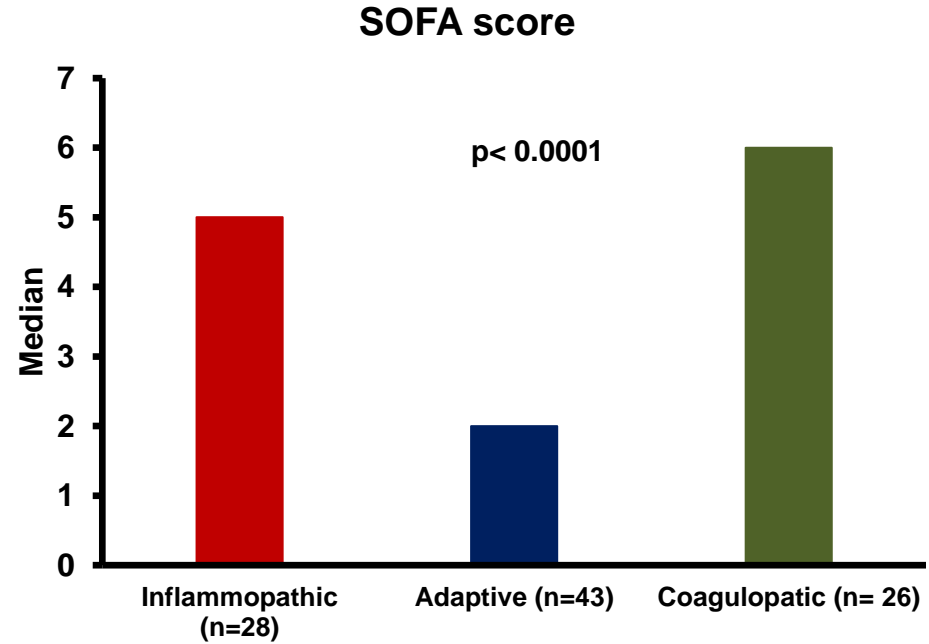
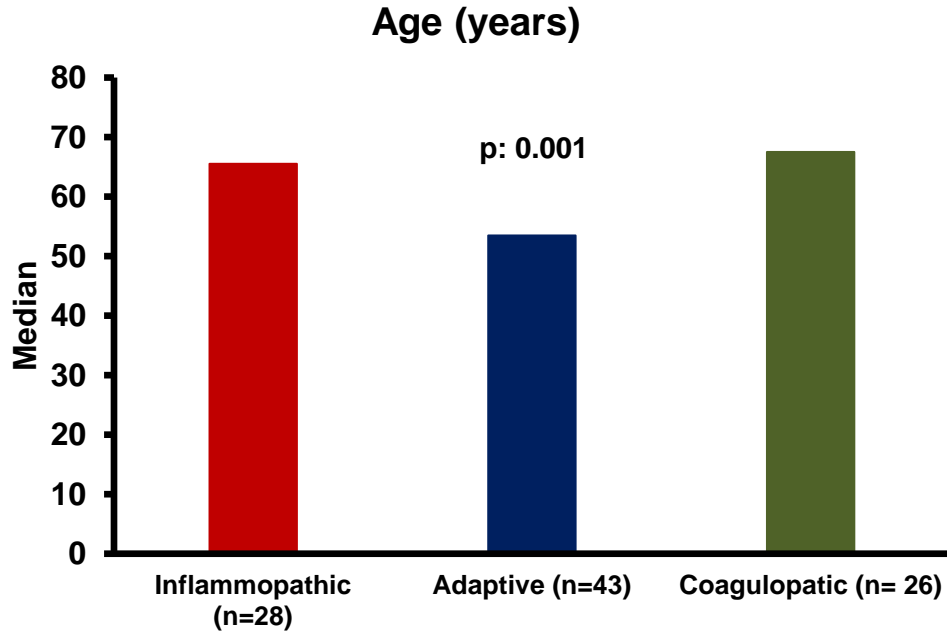
THE PATIENT POPULATION

	Survivors (n=81)	Non-survivors (n=16)	p
Age (years)	60.0 (50.8-70.3)	68.5 (62.8-84.3)	0.003
Male gender (%)	56 (69.1)	12 (75.0)	0.865
White blood cells	6,480 (5,145-9,622)	8,540 (5,542- 12,510)	0.275
Lymphocytes	1049.5 (759.7-1395.7)	613.8 (377.9-831.3)	<0.001
Platelets	214,000 (172,600- 260,800)	249,050 (180,750, 298,000)	0.176
D-dimers, ng/ml	850.0 (437.5-1947.5)	4480.0 (2440.0-13161.5)	<0.001
CRP, mg/l	79.1 (28.8-202.0)	224.8 (142.9-260.8)	0.002
IL-6, pg/ml	10.0 (10.0-59.0)	22.5 (10.0-135.0)	0.355
suPAR, ng/ml	4.80 [3.00, 6.00]	7.80 [5.50, 9.65]	0.002
Ferritin, ng/ml	633.0 (362.5-1324.0)	1407.0 (302.5-5033.5)	0.195
SOFA score	2 (1-6)	5 (4-6.3)	0.006
APACHE II	7.0 (4.0-9.0)	11.0 (8.0-13.5)	0.001
Hospital stay (days)	13.0 (11.0-20.0)	13.0 (8.8-17.3)	0.41
Mechan. Ventil. (%)	34 (42.0)	16 (100.0)	<0.001

APACHE: acute physiology and chronic health evaluation score
 CRP: C-reactive protein; IL: interleukin
 SOFA; sequential organ failure assessment
 suPAR: soluble urokinase plasminogen activator receptor

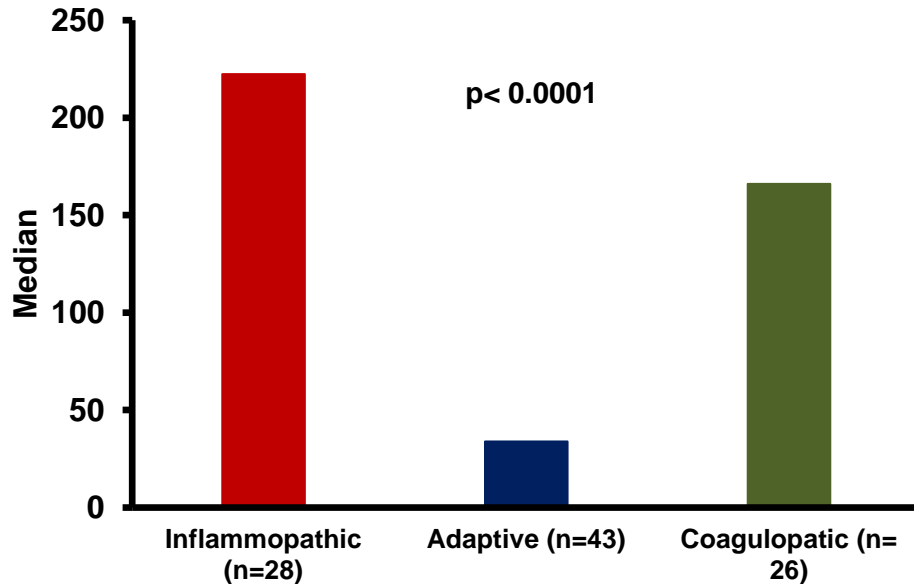
Values reported as medians and quartiles

TRANSLATION INTO DAILY ROUTINE

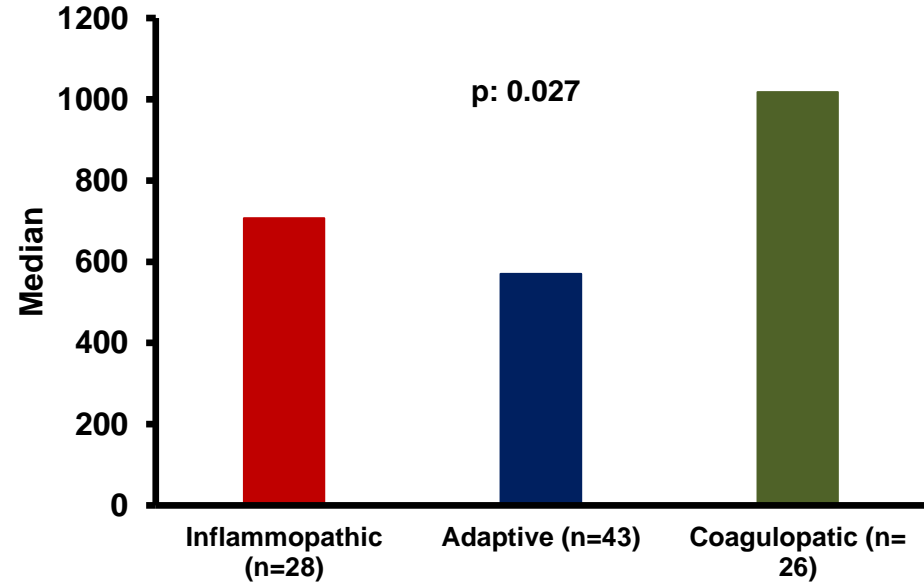


ASSOCIATION WITH INFLAMMATION

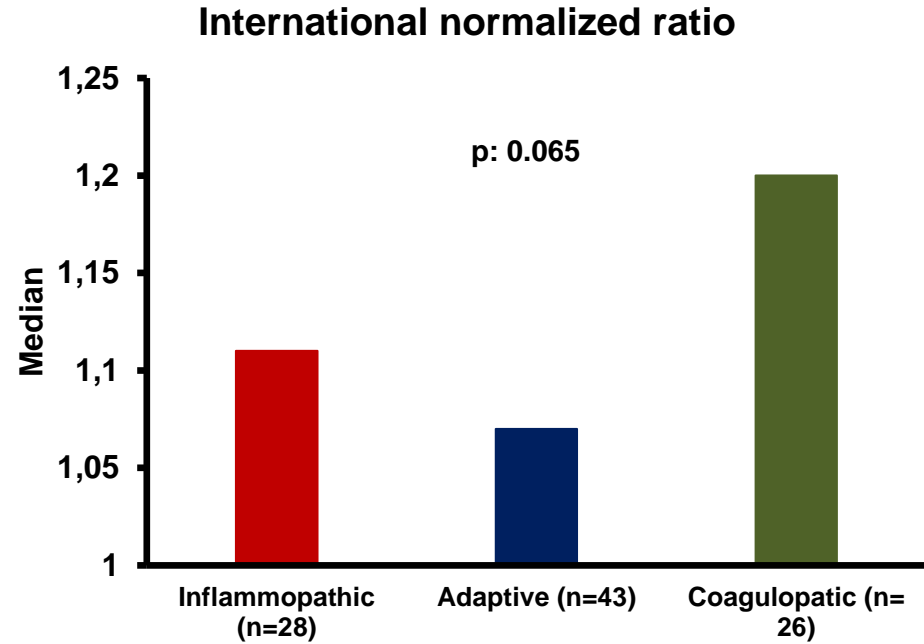
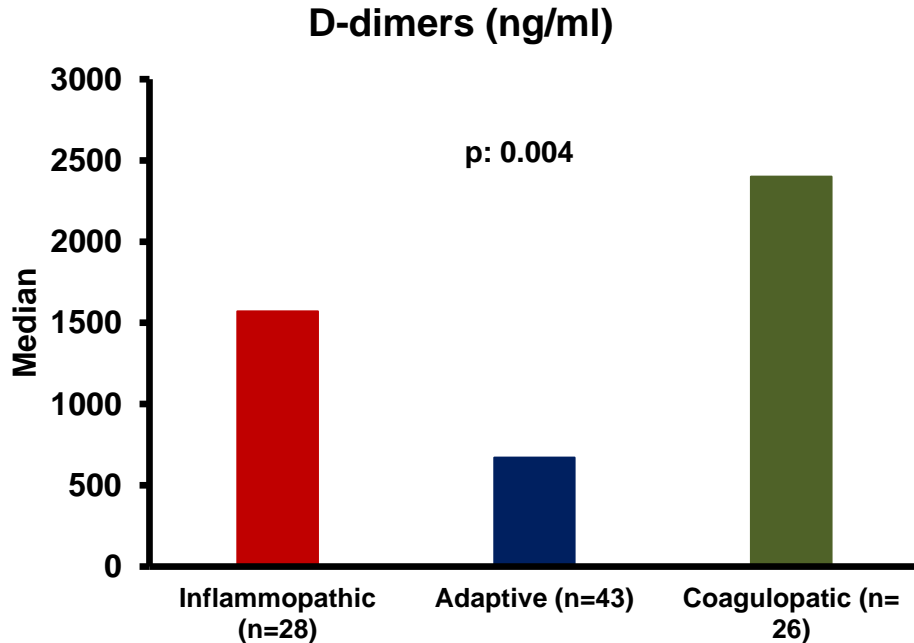
C-reactive protein (mg/l)



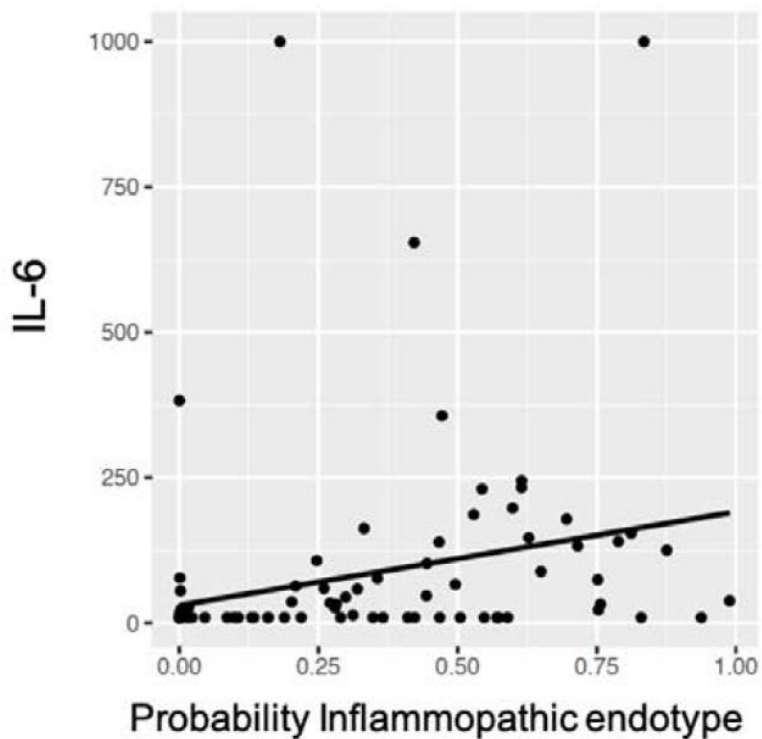
Ferritin (ng/ml)



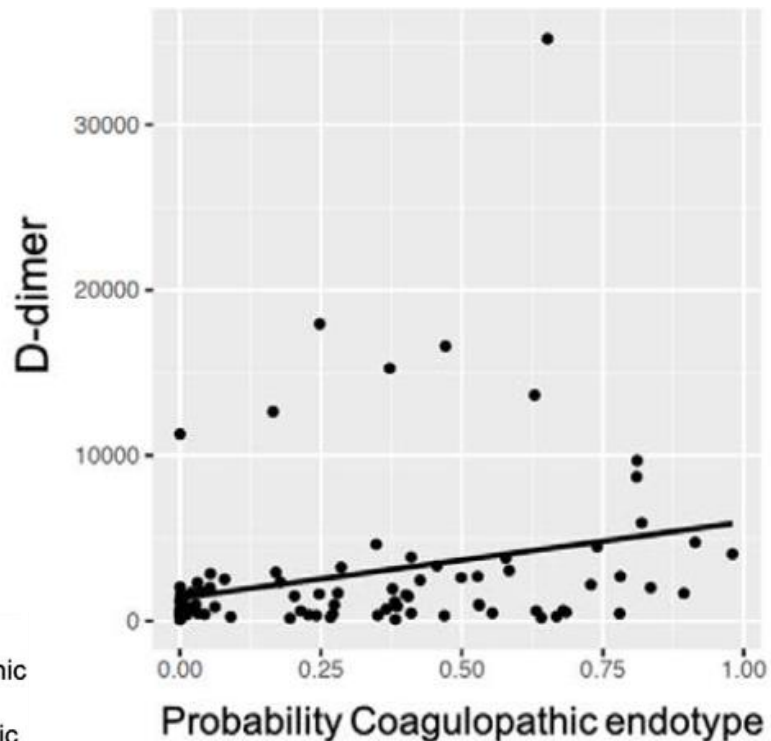
ASSOCIATION WITH COAGULATION



STATISTICAL PROBABILITY & BIOMARKERS

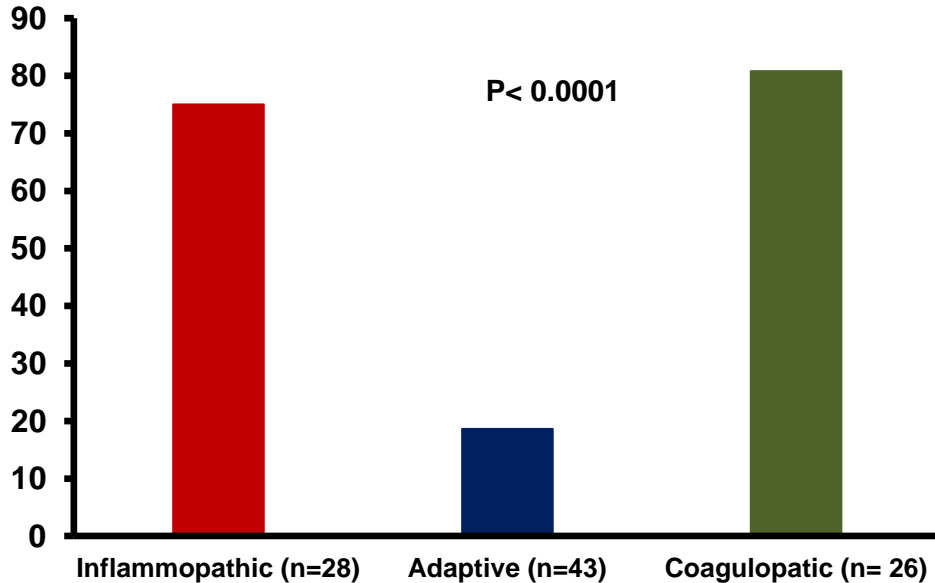


- Endotype
- Inflammopathic
 - Adaptive
 - Coagulopathic

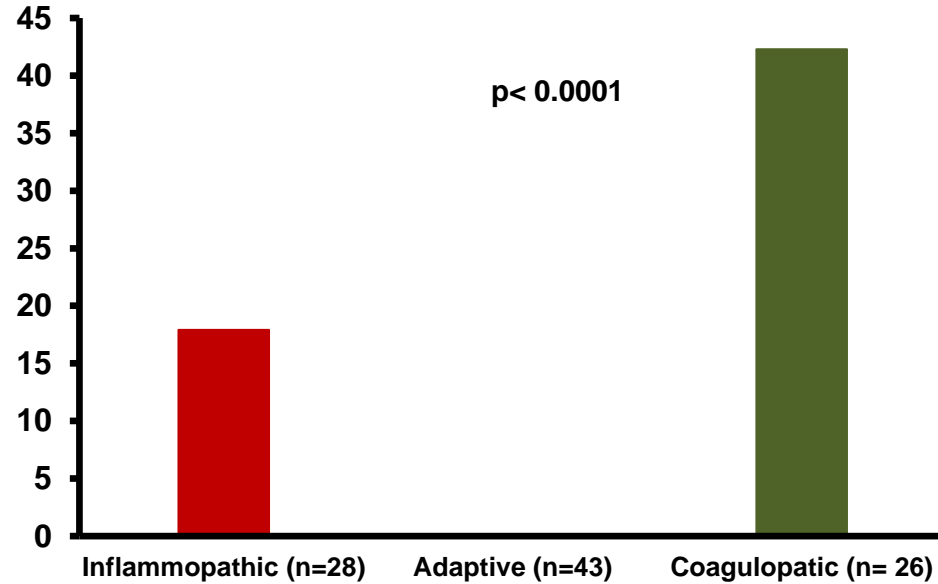


ASSOCIATION WITH OUTCOME

Mechanical ventilation (%)



28-day mortality (%)



CONCLUSIONS

- Heterogeneity of severe COVID-19
- Use a 33-mRNA read-out of the host

Inflammopathic endotype

- 5 genes
- Mortality ~20%
- ↑ CRP, IL-6
- Macrophage activation?

Coagulopathic endotype

- 11 genes
- Mortality >40%
- ↑ D-dimers
- Endothelial activation?

Adaptive endotype

- 17 genes
- Mortality <10%
- T-cell activation?

Validation of Inflammopathic, Adaptive, and Coagulopathic Sepsis Endotypes in Coronavirus Disease 2019

OBJECTIVES: Complex critical syndromes like sepsis and coronavirus disease 2019 may be composed of underlying "endotypes," which may respond differently to treatment. The aim of this study was to test whether a previously defined bacterial sepsis endotypes classifier recapitulates the same clinical and immunological endotypes in coronavirus disease 2019.

DESIGN: Prospective single-center observational cohort study.

SETTING: Patients were enrolled in Athens, Greece, and blood was shipped to Inflammatix (Burlingame, CA) for analysis.

PATIENTS: Adult patients within 24 hours of hospital admission with coronavirus disease 2019 confirmed by polymerase chain reaction and chest radiography.

INTERVENTIONS: None.

MEASUREMENTS AND MAIN RESULTS: We studied 97 patients with coronavirus disease 2019, of which 50 went on to severe respiratory failure (SRF) and 16 died. We applied a previously defined 33-messenger RNA classifier to assign endotype (Inflammopathic, Adaptive, or Coagulopathic) to each patient. We tested endotype status against other clinical parameters including laboratory values, severity scores, and outcomes. Patients were assigned as Inflammopathic (29%), Adaptive (44%), or Coagulopathic (27%), similar to our prior study in bacterial sepsis. Adaptive patients had lower rates of SRF and no deaths. Coagulopathic and Inflammopathic endotypes had 42% and 18% mortality rates, respectively. The Coagulopathic group showed highest D-dimers, and the Inflammopathic group showed highest C-reactive protein and interleukin-6 levels.

Timothy E. Sweeney, MD, PhD

Oliver Liesenfeld, MD*

James Wacker, MS*

Yudong D. He, PhD*

David Rawling, PhD*

Melissa Barwood, MS*

Sebrina Coyle, BA*

Urea Midzi, PhD*

Antigone Kotsaki, MD, PhD*

Aggeliki Kanarou, MD*

Konstantinos Leventogiannis, MD*

Ioanna Kontogiorgos, MD*

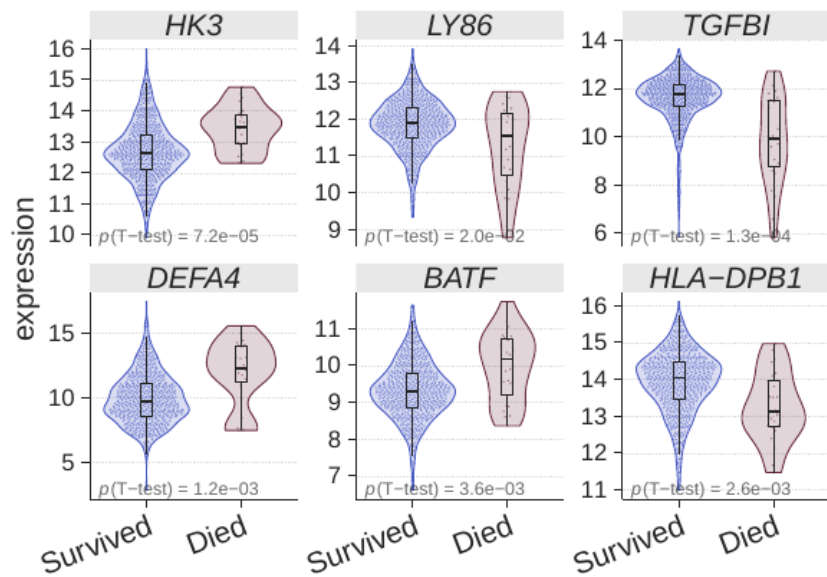
Evangelos J. Giamarellos-

Bourboulis, MD, PhD*

THE FUTURE: HOST SIGNATURE FOR PROGNOSIS

(Buturovic L, et al. medRxiv doi.org/10.1101/2020.12.07.20230235)

- Signature from 6 gene copies generated in non-COVID-19 patients
- Application in the same cohort of 97 patients



THE NEW GENOMIC SCORE FOR PROGNOSIS

(Buturovic L, et al. medRxiv doi.org/10.1101/2020.12.07.20230235)

